



## ABSTRACT

**Objectives:** Photodynamic therapy (PDT) was first used to eliminate microorganisms over 100 years ago, and is known to require an ample oxygen supply. Infective biofilms grown in periodontal pockets are oxygen-starved and difficult to characterize. To evaluate the potential “in-pocket” efficacy of this re-emerging therapy for periodontal infection control, a novel methodological procedure of small-space biofilm growth and characterization must be developed.

**Methods:** We are now testing the utility of Multiple Attenuated Internal Reflection – Infrared Spectroscopy, using germanium miniature prisms and commercially pure titanium test pieces, anaerobically incubated for 24 hours with *Fusobacterium nucleatum* to allow for biofilm formation.

**Results:** Specimens dried and mounted upon the appropriate apparatus confirmed biofilm growth by using MAIR-IR spectroscopy and collecting the spectral data output. Biofilm growth is corroborated using SEM analysis and a colony-forming unit (CFU) count technique termed- “drop spot counting.”

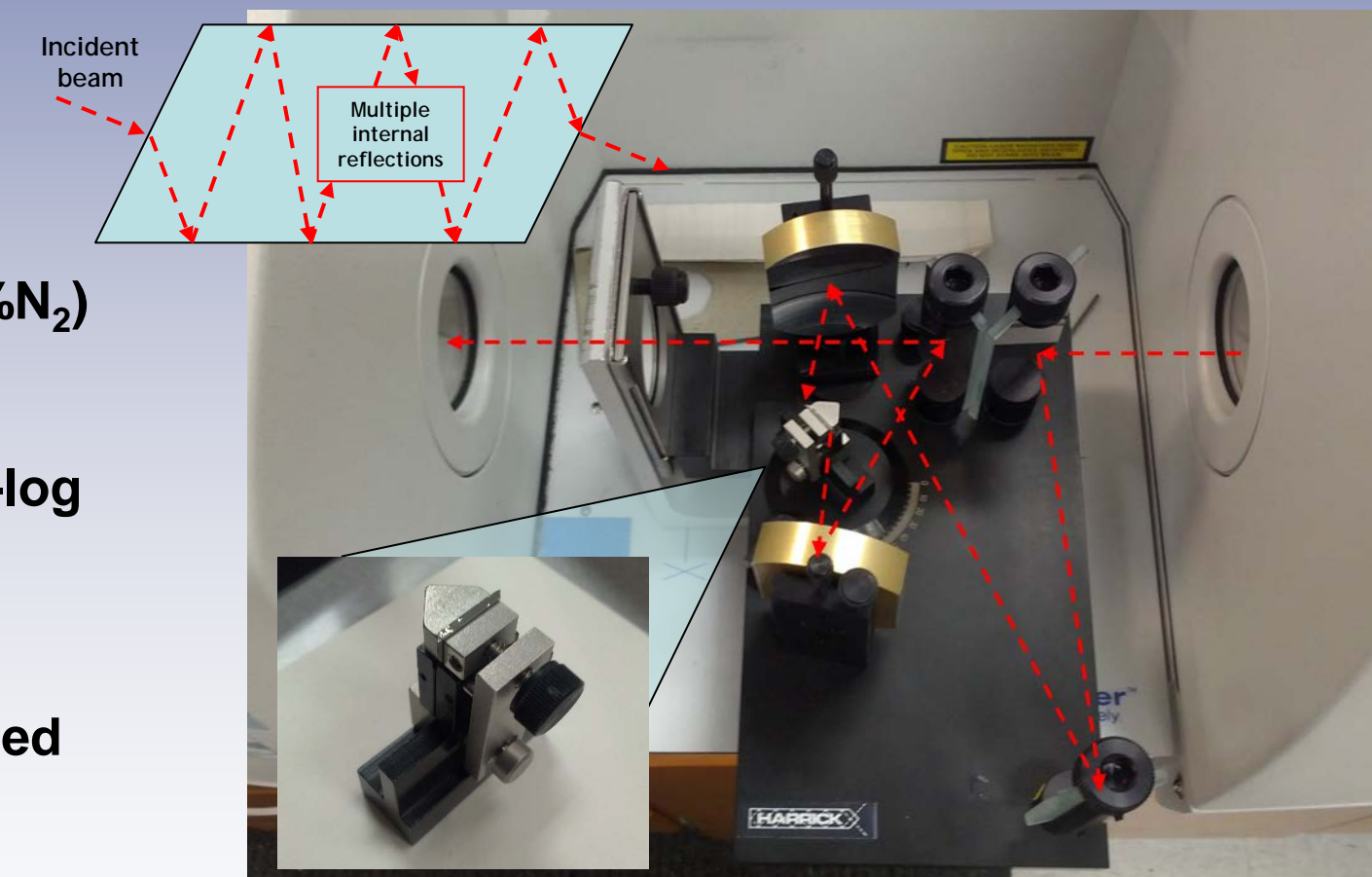
**Conclusion:** The current data suggest that this method has the potential, in combination with other methodologies, to serve as a reference point for PDT efficacy studies.

## INTRODUCTION

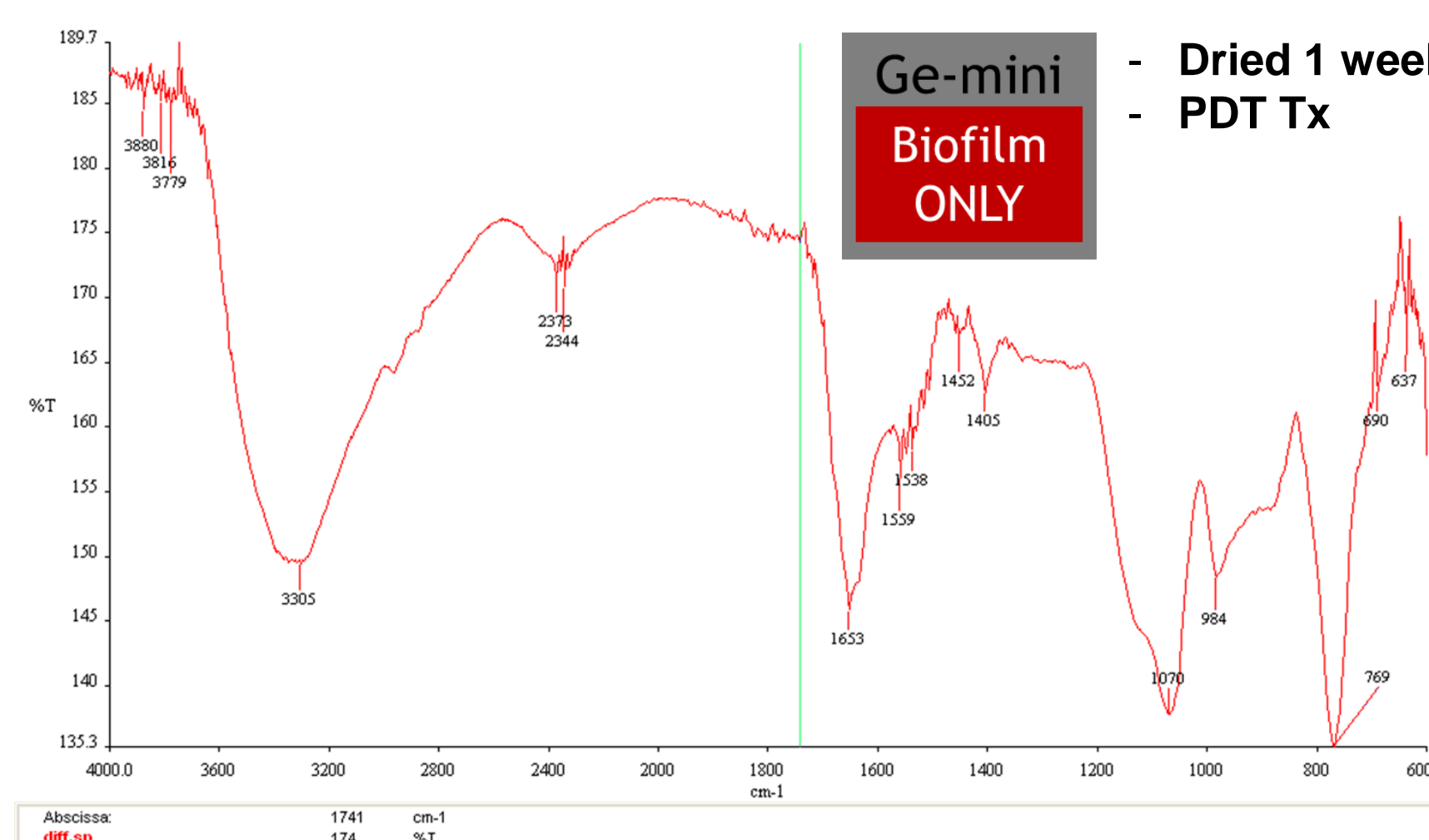
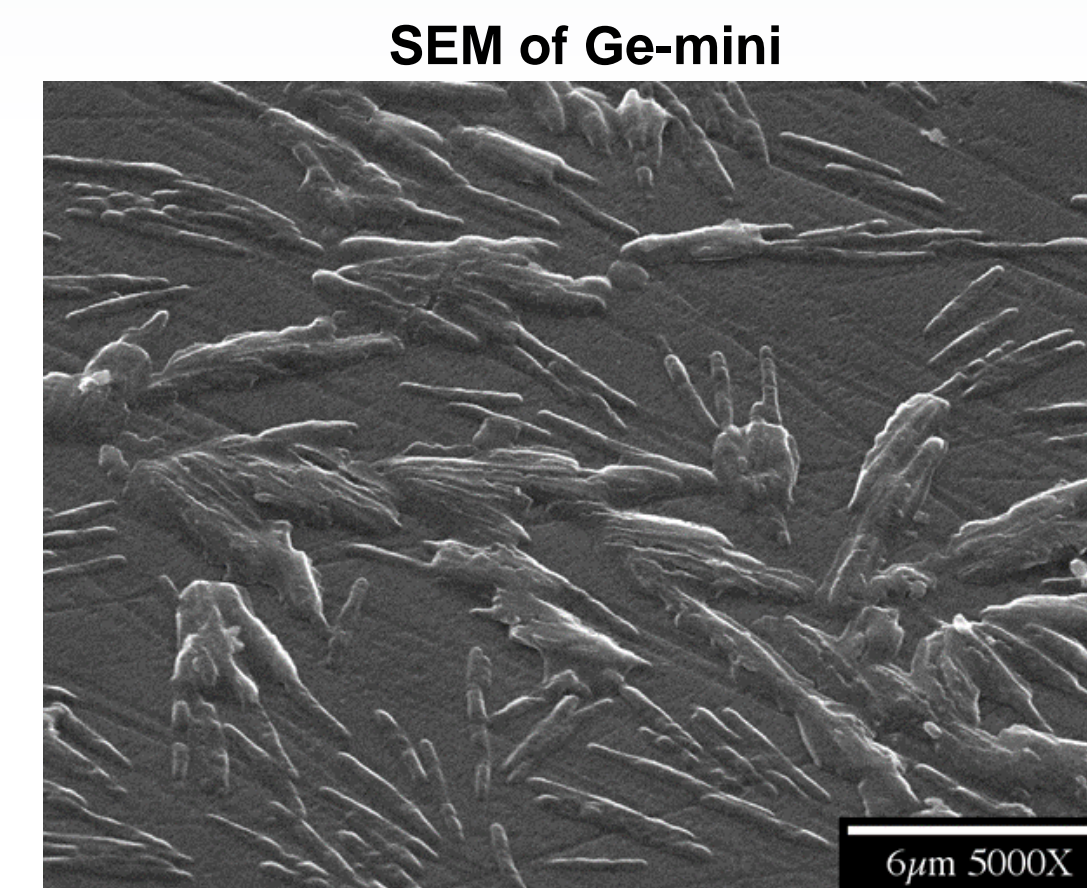
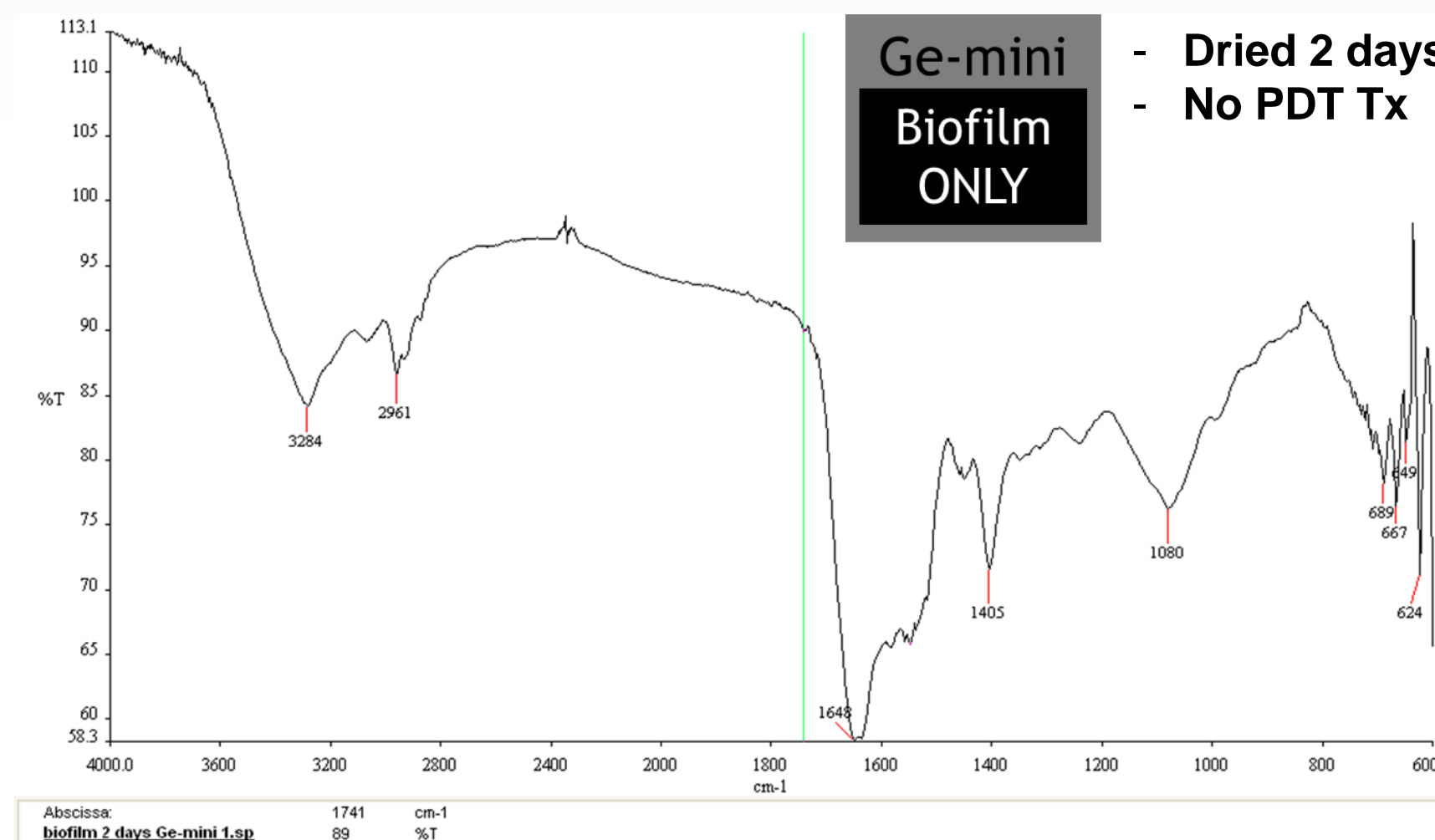
- Common oral pathogen grown on miniature germanium prisms to characterize growth via Multiple Attenuated Internal Reflection Infrared Spectroscopy (MAIR-IR)
- Specimens dried and either subjected to a photosensitizer(PS) + PDT, or no treatment
- CFU counts<sup>b</sup> conducted to ascertain treatment effectiveness
- SEM images collected to inspect surface for growth

## MATERIALS and METHODS

- *Fusobacterium nucleatum* (ATCC 25586) maintained at 37degC in tryptic soy broth (DIFCO) containing:
  - 0.1% L-cysteine + 0.5% yeast extract
  - anaerobic condition (10%CO<sub>2</sub>; 10%H<sub>2</sub>; 80%N<sub>2</sub>)
- Number of bacteria adjusted to OD 0.3-0.4 at 600nm after bacterial growth reached the mid-log phase
- 2mL of bacterial solution applied to 6-well cell culture plates (Costar) with specimens, wrapped with cellophane



## RESULTS



### Colony Forming Units (viable bacteria)

	specimen	Laser time (sec)	CFU*
1	Ti	untreated	208,000/ml
2	Ge	untreated	192,000/ml
3	Ge	167	90,000/ml
4	Ge	167	10,000/ml
5	Ge	167	330,000/ml
6	Ti	167	100/ml
7	Ti	333	1,670/ml
8	Ti	333	67/ml
9	Ti	667	0
10	Ti	667	0

## CONCLUSIONS

- Between 2 days and 7 days, there is considerable biofilm maturation shown by a substantial increase of “slime layer” (polysaccharides, ~1080cm<sup>-1</sup>) and the prevalence of multiple hydroxide and hydrocarbon bands
- Although PDT was killing, as suggested by CFU counts<sup>b</sup>, there was not much removal of the biofilm
- Literature supports PDT as an effective antimicrobial therapy in the oral cavity<sup>a,d</sup>
- This effectiveness is limited by the very nature of biofilms and their inherent antimicrobial resistance in the form of an exopolymer exudate<sup>c</sup>
- Despite PDT treatment, this biofilm will likely reestablish itself and continue to thrive

## REFERENCES

<sup>a</sup>Fontana, C. R., A. D. Abernethy, S. Som, K. Ruggiero, S. Doucette, R. C. Marcantonio, C. I. Boussios, R. Kent, J. M. Goodson, A. C. Tanner, and N. S. Soukos. "The Antibacterial Effect of Photodynamic Therapy in Dental Plaque-Derived Biofilms." *J Periodontal Res* 44.6 (2009): 751-9. Print.

<sup>b</sup>Honma, K., E. Mishima, and A. Sharma. "Role of Tannerella Forsythia Nanh Sialidase in Epithelial Cell Attachment." *Infect Immun* 79.1 (2011): 393-401. Print.

<sup>c</sup>Mang, T. S., D. P. Tayal, and R. Baier. "Photodynamic Therapy as an Alternative Treatment for Disinfection of Bacteria in Oral Biofilms." *Lasers Surg Med* 44.7 (2012): 588-96. Print.

<sup>d</sup>Soukos, N. S., and J. M. Goodson. "Photodynamic Therapy in the Control of Oral Biofilms." *Periodontol* 2000 55.1 (2011): 143-66. Print.

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